

硝普钠及其光解产物对日本晴水稻幼苗生长和 5 种激素标记基因表达的影响

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摘要:以粳稻品种日本晴水稻为材料,检测硝普钠及其光解产物 KNO_2 、 $\text{K}_4\text{Fe}(\text{CN})_6$ 对幼苗生长的影响,并采用 qRT-PCR 技术检测了 5 种植物激素标记基因在经上述处理后在幼苗根中的表达水平.结果表明 SNP 能够显著抑制水稻幼苗的根长和株高;SNP 对根生长的抑制主要是通过其光解产物 $\text{K}_4\text{Fe}(\text{CN})_6$ 实现的. SNP 和 $\text{K}_4\text{Fe}(\text{CN})_6$ 处理都能够抑制生长素、细胞分裂素、脱落酸和赤霉素 4 种激素标志基因在水稻根中的表达,但是 SNP 处理可以抑制一氧化氮标志基因 OsNOA1 的表达,而 $\text{K}_4\text{Fe}(\text{CN})_6$ 则上调该基因的表达.

关键词:硝普钠;水稻;幼苗;植物激素;基因表达

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在高等植物中,根是重要的营养器官之一,具有吸收、储藏和运输营养物质的功能.更重要的是,根部产生的植物激素影响着地上部分的生长和发育.以往的研究表明,多种植物激素参与调控根的生长发育,其中生长素、细胞分裂素及赤霉素均能促进根的生长^[1].近期的研究发现,气体信号分子一氧化氮(nitric oxide, NO)也参与植物根的发育调节过程^[2~3],其作用方式涉及植物激素信号传导网络,但是机制尚不清楚.因此,对植物根生长发育与 NO 等激素信号关系的研究有待深入.

水稻是重要的粮食作物和模式植物,已有的研究表明 NO 影响水稻根的形态建成,包括水稻侧根的形成^[4].硝普钠(sodium nitroprusside, SNP)作为研究常用的 NO 供体,经过光解后会产生 3 种产物即 NO, KNO_2 和 $\text{K}_4\text{Fe}(\text{CN})_6$,尽管已有 SNP 对水稻生长发育影响的检测^[5~7],但是 SNP 及其光解产物对水稻生长发育的系统研究尚未见报道.为此本文以粳稻品种日本晴为材料,以外源 NO 供体 SNP 以及光解产物 KNO_2 , $\text{K}_4\text{Fe}(\text{CN})_6$ 处理幼苗,并采用 qRT-PCR 检测在上述处理条件下水稻植物激素标记基因的表达水平,旨在检测其对水稻幼苗生长的影响,并明确 SNP 处理对生长素、细胞分裂素等植物激素水平的影响,为进一步鉴定 NO 在水稻生长发育中的功能奠定基础.

1 材料和方法

1.1 供试材料

水稻粳稻品种日本晴(*Oryza sativa L. japonica. cv. Nipponbare*)由本实验室保存.

1.2 供试试剂

硝普钠, $\text{K}_4\text{Fe}(\text{CN})_6$, KNO_2 购于北京鼎国生物有限公司;总 RNA 提取试剂 TRNzol, DNaseI 酶, Prime-Script RT Master Mix 购于大连宝生物有限公司;SYBR Green Master Mix 购于南京诺唯赞生物技术有限公司.

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1.3 方法

1.3.1 水稻幼苗的培养和处理

取水稻种子适量用75%的酒精消毒,无菌水冲洗,室温浸泡1 h后,置于人工气候箱28℃黑暗培养催芽,待种子发芽后转移至Yoshida完全营养液^[8]进行水培,培养温度28℃,光照强度为16 000 lx,光照时间为12 h/d.外源NO处理采用SNP添加到Yoshida完全营养液中,终浓度分别为0、0.02、0.04、0.06、0.08 mM,每4 d更换一次营养液,培养12 d取样,测量不同处理组水稻的根长和株高,每组样品重复3次.

1.3.2 总RNA的提取

取生长至12 d的水稻幼苗根,液氮研磨后,按照TRNzol说明书的要求提取总RNA,并采用超微量光度计检测RNA样品浓度及质量.

1.3.3 反转录

按照宝生物反转录试剂盒说明书的要求进行反转录,制备cDNA反应体系如下:5×PrimeScript RT Master Mix(2 μL)、Total RNA(≤500 ng)、RNase Free H₂O(补至10 μL).将上述反应体系短暂离心混匀后,在37℃保温15 min,然后85℃5 s终止反应.

1.3.4 实时定量PCR

用以上制备的cDNA为模板,内参基因选用*OsAct1*,其他检测基因引物见表1.按照SYBR Green Master Mix说明书进行配制20 μL体系:2×SYBR Green Master Mix(10 μL)、上游引物(0.4 μL)、下游引物(0.4 μL)、ROX Reference Dye II (50×)(0.4 μL)、cDNA(2 μL)和RNase Free H₂O(6.8 μL).采用两步法进行PCR扩增:95℃预变性,5 min;PCR反应95℃,10 s;然后60℃,30 s,并收集数据.设置未处理水稻为对照组,每个样品平行重复3次.

表1 实时定量PCR引物一览表

引物序号	引物序列 (5'-3')	基因名称	基因特征
P1	TCTTCCAGCCTTCCTTCA	<i>OsAct1</i>	肌动蛋白基因 ^[9]
P2	ATCCACGTCGCACTTCAT		
P3	ATAATCCTCGCCACAGGCTACA	<i>OsYUCCA1</i>	IAA 标记基因 ^[10]
P4	GCAGTGCAGACAGAAAGAAAA		
P5	TACGAGTGCTGCTTCCTCTGGG	<i>OsIPT</i>	CTK 标记基因 ^[11]
P6	AGATGCCCTGGAGTAGTCGGT		
P7	ATCGGCTGGAGATGAAGAGGG	<i>OsGA20ox1</i>	GA 标记基因 ^[12]
P8	GCGGCTCATCTCGTGCCAGT		
P9	ACTGTGTCGAGCCTGTG	<i>OsNCED2</i>	ABA 标记基因 ^[13]
P10	GGAAGGTCAGATTCGCATAG		
P11	TGAAGGATGGGTTGGTCTG	<i>OsNOA1</i>	NO 标记基因 ^[14]
P12	ACCCACCCGAGACCAGAAA		

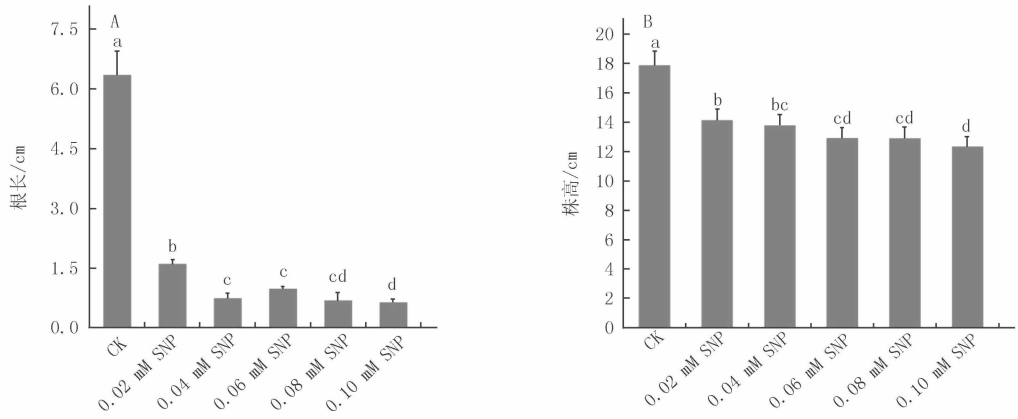
1.4 数据分析

采用DPS数据处理软件对水稻根长和株高数据进行方差分析,实时定量PCR的数据采用 $2^{-\Delta\Delta C_t}$ 法进行处理分析.数据分析结果用Origin 8.5软件绘制图表.

2 结果与分析

2.1 SNP对水稻幼苗生长的影响

采用不同浓度的SNP处理水稻幼苗,统计根长和株高,发现与未处理的水稻相比,不同浓度的SNP对幼苗的根和地上部分的生长均有抑制作用(图1).结果显示,随着SNP处理浓度的升高,水稻的根长和株高都明显的降低,统计学分析显示SNP对地上部分生长的抑制程度不如对根的抑制显著.与对照相比,随着SNP浓度的增加,根长降低幅度为88%~90%.



“a, b, c, d”代表 $P < 0.05$ 显著分析

图1 不同浓度SNP对水稻幼苗根长(A)和株高(B)的影响

2.2 SNP及其光解产物对水稻幼苗生长的抑制效应分析

为了解SNP抑制根生长的作用机制,进一步采用SNP及其光解产物 KNO_2 和 $\text{K}_4\text{Fe}(\text{CN})_6$ 分别处理幼苗,与对照根长进行比较,结果显示,SNP和 $\text{K}_4\text{Fe}(\text{CN})_6$ 处理后,根的长度与对照相比显著降低,SNP处理根长略微短于 $\text{K}_4\text{Fe}(\text{CN})_6$ 处理,而 KNO_2 处理后,水稻幼根长度与对照差别不大,甚至略有增加(图2,图3A)。此外,对株高的统计也显示出类似的结果(图3B),表明SNP对根生长的抑制效应中, $\text{K}_4\text{Fe}(\text{CN})_6$ 发挥主要作用。

2.3 SNP及其光解产物对激素标记基因在水稻幼苗根中表达的影响

分别选择5种植物激素生长素(IAA)、细胞分裂素(CTK)、赤霉素(GA)、脱落酸(ABA)和NO的标记基因 OsYUCCA1 , OsIPT , OsGA20ox1 , OsNCED2 和 OsNOA1 ,采用qRT-PCR检测上述激素标记基因在经SNP及其光解产物处理后的水稻根中表达特征,结果显示,SNP处理后,5种基因表达都下调;而 $\text{K}_4\text{Fe}(\text{CN})_6$ 处理后,除 OsNOA1 基因表达上调外(图4E),其余4种基因表达都下调;另外 KNO_2 处理后,除了 OsIPT (图4B)和 OsNCED2 (图4C)这两种基因表达与对照组没有显著差异外,其他3种标记基因表达水平都显著上调。

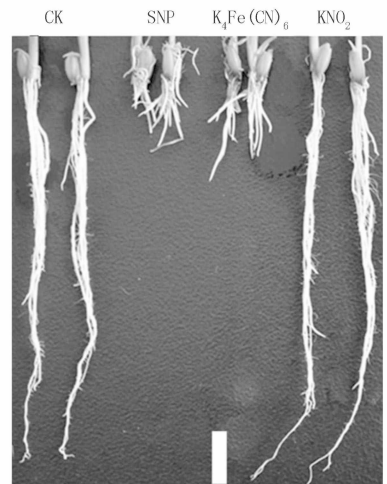


图2 SNP及其光解产物对水稻幼苗根长的影响
处理浓度为0.06 mM, Bar=1 cm

3 讨论

NO已被证明是普遍存在于动植物中的气体信号分子。已有的报道表明,NO参与植物多种生长发育过程的调节,例如植物根的形态建成^[15],植物细胞的程序性死亡^[16],气孔的关闭^[17]等。本研究用SNP处理水稻幼苗,结果显示水稻幼苗根系伸长生长被抑制,根的形态与处理前相比也发生明显的改变,侧根呈现短粗特征。SNP处理后,水稻株高也明显的降低,可能是由于SNP对根生长的抑制,影响了根对营养的吸收,可能还引起植物激素水平降低,导致水稻株高的降低。

由于SNP光解时生成 KNO_2 , $\text{K}_4\text{Fe}(\text{CN})_6$ 和NO 3种物质,为此,本研究采用SNP及其光解产物分别处理水稻进一步研究SNP的作用机制。结果显示,SNP与 $\text{K}_4\text{Fe}(\text{CN})_6$ 都能够抑制水稻根的生长,且两种处理水稻幼根的表型相似,提示在上述条件下SNP对根生长的抑制效应是NO和 $\text{K}_4\text{Fe}(\text{CN})_6$ 共同作用的结果,且以 $\text{K}_4\text{Fe}(\text{CN})_6$ 为主。

已有研究表明NO与多种植物激素信号途径相关,比如通过参与IAA^[18-19],CTK^[20],ABA^[21],乙

烯^[22],水杨酸^[23]等植物激素之间的信号转导网络来调节植物生理过程. 因此在 SNP 及其光解产物处理的条件下,水稻幼苗生长受抑制的过程中,其他植物激素水平是否发生变化值得研究. 为此,本研究选取了 5 种水稻植物激素的标记基因,进行 qRT-PCR 检测,发现 SNP 和 $K_4Fe(CN)_6$ 处理都能下调 IAA,CTK 和 GA 标记基因在根中的表达水平,提示可能会因此导致上述激素含量的降低,说明外源 NO 和 $K_4Fe(CN)_6$ 可能通过降低水稻幼苗根中 IAA,CTK,GA 的水平,达到对根生长的抑制效应,在功能上与上述激素发生拮抗作用. SNP 处理的效应显著于 $K_4Fe(CN)_6$ 处理,也进一步证明 SNP 的复合效应. 另外,值得注意的是, KNO_2 和 $K_4Fe(CN)_6$ 处理都明显提高了 *OsNOA1* 基因的表达水平,推测由此导致水稻内源 NO 水平的提高,提示 SNP 不仅提供外源 NO,也影响水稻内源 NO 水平. 本研究为探究 NO 在抑制水稻根生长中的作用机制以及 与 IAA,CTK 等 5 种植物激素的关系提供了线索,为揭示水稻幼苗根发育过程中复杂的信号网络奠定理论基础.

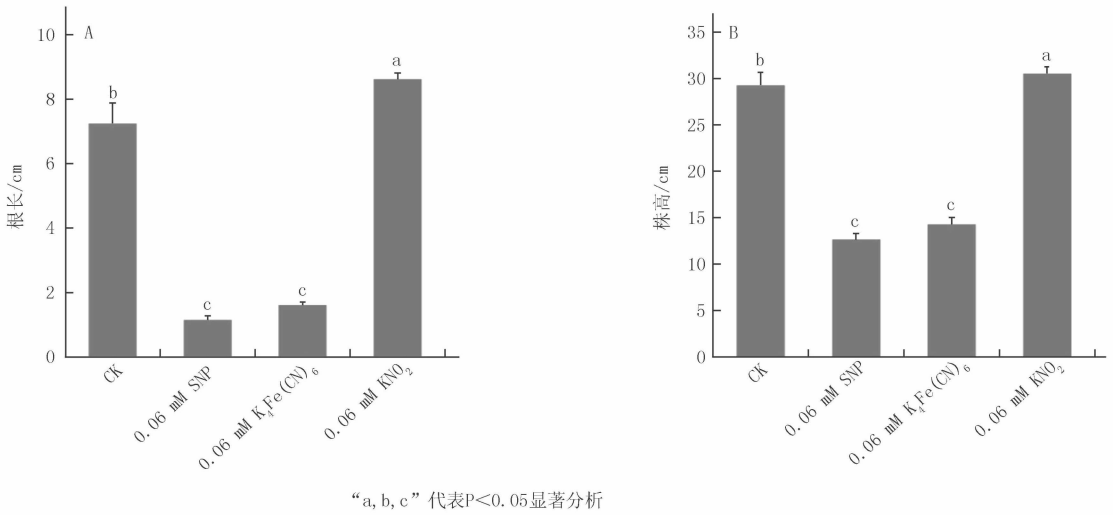
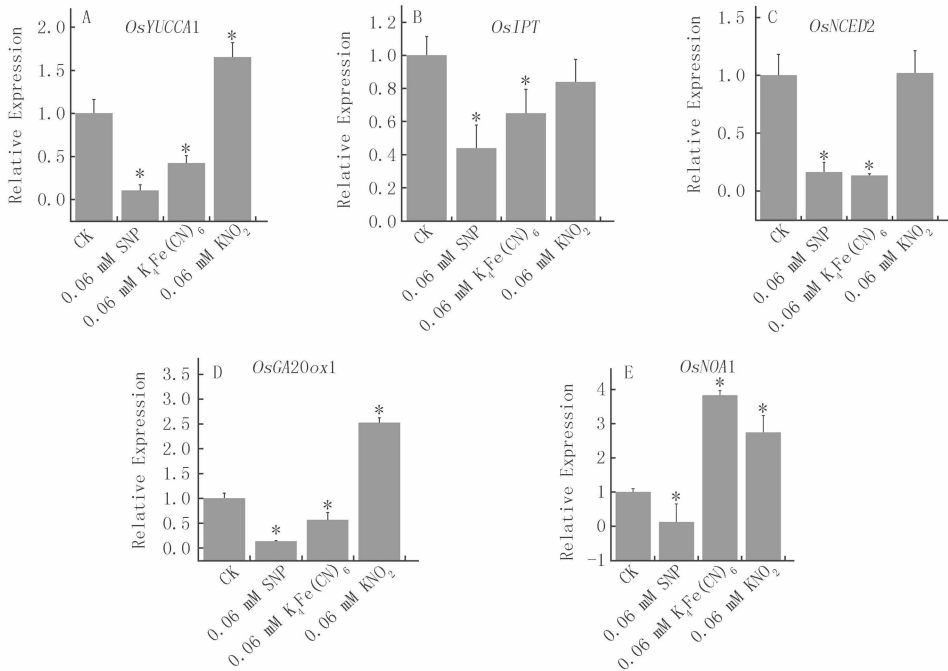


图3 SNP 及其光解产物对水稻幼苗根长(A)和株高(B)的统计学分析



*代表 $P < 0.05$ 显著分析

图4 SNP 及其光解产物对水稻5种植物激素标志基因在水稻根中表达的影响

参 考 文 献

- [1] Druege U, Franken P, Hajirezaei M R. Plant Hormone Homeostasis, Signaling, and Function during Adventitious Root Formation in Cuttings[J]. *Frontiers in Plant Science*, 2016, 7:381.
- [2] Chen Y H, Kao C H. Calcium is involved in nitric oxide-and auxin-induced lateral root formation in rice[J]. *Protoplasma*, 2012, 249(1): 187-195.
- [3] Xiong J, Lu H, Lu K, et al. Cadmium decreases crown root number by decreasing endogenous nitric oxide, which is indispensable for crown root primordia initiation in rice seedlings[J]. *Planta*, 2009, 230(4): 599-610.
- [4] Chen Y H, Chao Y Y, Hsu Y Y, et al. Heme oxygenase is involved in nitric oxide-and auxin-induced lateral root formation in rice[J]. *Plant Cell Reports*, 2012, 31(6): 1085-1091.
- [5] Liu H Y, Yu X, Cui D Y, et al. The role of water channel proteins and nitric oxide signaling in rice seed germination[J]. *Cell Research*, 2007, 17(7): 638-649.
- [6] Chen Y H, Kao C H. Calcium is involved in nitric oxide-and auxin-induced lateral root formation in rice[J]. *Protoplasma*, 2012, 249(1): 187-195.
- [7] Xiong J, Lu H, Lu K, et al. Cadmium decreases crown root number by decreasing endogenous nitric oxide, which is indispensable for crown root primordia initiation in rice seedlings[J]. *Planta*, 2009, 230(4): 599-610.
- [8] Yoshida S, Forno D A, Cock J H, et al. Laboratory manual for physiological studies of rice[M]. 3rd ed. Manila: Int Rice Res Inst, 1972.
- [9] Shin R, Schachtman D P. Hydrogen peroxide mediates plant root cell response to nutrient deprivation[J]. *Proceedings of the National Academy of Sciences of the United States of America*, 2004, 101(23): 8827-8832.
- [10] Yamamoto Y, Kamiya N, Morinaka Y, et al. Auxin biosynthesis by the YUCCA genes in rice[J]. *Plant Physiology*, 2007, 143(3): 1362-1371.
- [11] Sakamoto T, Sakakibara H, Kojima M, et al. Ectopic expression of KNOTTED1-like homeobox protein induces expression of cytokinin biosynthesis genes in rice[J]. *Plant Physiology*, 2006, 142(1): 54-62.
- [12] Oikawa T, Koshioka M, Kojima K, et al. A role of OsGA20ox1, encoding an isoform of gibberellin 20-oxidase, for regulation of plant stature in rice[J]. *Plant Molecular Biology*, 2004, 55(5): 687-700.
- [13] Zhu G, Ye N, Zhang J. Glucose-induced delay of seed germination in rice is mediated by the suppression of ABA catabolism rather than an enhancement of ABA biosynthesis[J]. *Plant and Cell Physiology*, 2009, 50(3): 644-651.
- [14] Qiao W, Xiao S, Yu L, et al. Expression of a rice gene OsNOA1 re-establishes nitric oxide synthesis and stress-related gene expression for salt tolerance in Arabidopsis nitric oxide-associated 1 mutant Atnoa1[J]. *Environmental and Experimental Botany*, 2009, 65(1): 90-98.
- [15] Fernández-Marcos M, Sanz L, Lewis D R, et al. Nitric oxide causes root apical meristem defects and growth inhibition while reducing PIN-FORMED 1 (PIN1)-dependent acropetal auxin transport[J]. *Proceedings of the National Academy of Sciences*, 2011, 108(45): 18506-18511.
- [16] Ye Y, Li Z, Xing D. Sorting out the role of nitric oxide in cadmium-induced Arabidopsis thaliana programmed cell death[J]. *Plant Signaling & Behavior*, 2012, 7(11): 1493-1494.
- [17] Zhao X, Qiao X, Yuan J, et al. Nitric oxide inhibits blue light-induced stomatal opening by regulating the K⁺ influx in guard cells[J]. *Plant Science*, 2012, 184: 29-35.
- [18] Petó A, Lehotai N, Lozano-Juste J, et al. Involvement of nitric oxide and auxin in signal transduction of copper-induced morphological responses in Arabidopsis seedlings[J]. *Annals of Botany*, 2011, 108(3): 449-457.
- [19] Schlicht M, Ludwig - Müller J, Burbach C, et al. Indole - 3 - butyric acid induces lateral root formation via peroxisome - derived indole - 3 - acetic acid and nitric oxide[J]. *New Phytologist*, 2013, 200(2): 473-482.
- [20] Liu W Z, Kong D D, Gu X X, et al. Cytokinins can act as suppressors of nitric oxide in Arabidopsis[J]. *Proceedings of the National Academy of Sciences*, 2013, 110(4): 1548-1553.
- [21] Hancock J T, Neill S J, Wilson I D. Nitric oxide and ABA in the control of plant function[J]. *Plant Science*, 2011, 181(5): 555-559.
- [22] Lin Y, Yang L, Paul M, et al. Ethylene promotes germination of Arabidopsis seed under salinity by decreasing reactive oxygen species: evidence for the involvement of nitric oxide simulated by sodium nitroprusside[J]. *Plant Physiology and Biochemistry*, 2013, 73: 211-218.
- [23] Zottini M, Costa A, De Michele R, et al. Salicylic acid activates nitric oxide synthesis in Arabidopsis[J]. *Journal of Experimental Botany*, 2007, 58(6): 1397-1405.

- [21] PHILLIPS T W, WIKING A J, ATKINSON T II, et al. Synergism of turpentine and ethanol as attractants for certain pine-infesting beetles (Coleoptera)[J]. *Environmental Entomology*, 1988, 17(3): 456-462.
- [22] NEIME M E, KEENA M A, ZHANG J R, et al. Attraction of *Anoplophora glabripennis* to male-produced pheromone and plant volatiles[J]. *Environmental Entomology*, 2009, 38(6): 1745-1755.

Extraction and Identification of Host-Plant Volatiles of *Acer mono* and EAG Responses of *Anoplophora nobilis* to the Primary Compounds of *A. mono* Volatiles

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Abstract: To explore the most appropriate hosts of nutrient supplement and oviposition for *Anoplophora nobilis* adults in Yili Prefecture, the volatiles composition of *Acer mono* were analyzed with GC-MS, and the electrophysiological activities of adults towards main volatiles were tested by EAG responses. The results showed that *A. nobilis* preferred fresh twigs of *A. mono* for diet supplement and carve twigs to oviposit in mixed plant area. Thirty-four volatiles were identified from shoots and leaves of *A. mono*, and terpenoids and esters were the major components, with the relative concentration up to 70.49% and 23.94%, respectively. (*Z*)- β -ocimene, sabinene, β -caryophyllene, α -phellandrene and cis-3-Hexenyl acetate represented a higher proportion in all components, with the relative concentration of 21.07%, 19.59%, 7.35%, 13.58% and 12.99%, respectively. With the concentration of 10 mg · mL⁻¹, the EAG responses of female adults to β -caryophyllene was significantly higher than the other volatile compounds, and the male showed the most intensively EAG responses to ethyl acetate and α -pinene. The EAG responses of male adults to ethyl acetate and α -pinene were significantly stronger than female, while the female adults had more strongly EAG responses to β -caryophyllene and cis-3-Hexenyl-2-methyl butyrate compared to males. This study will contribute to further research of chemical fingerprinting relating to host location of *A. nobilis*, therefore have a certain practical significance to development plant source attractants.

Keywords: *Anoplophora nobilis*; host selection; plant volatiles; electroantennogram responses; olfactory communication

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Effects of SNP and Its Photolysis Products on the Seedlings Growth of Rice and Expression of Marker Genes for Five Plant Hormones

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Abstract: Taking japonica cultivar Nipponbare rice as materials, the effects of sodium nitroprusside and its photolysis products KNO₂、K₄Fe(CN)₆ on the rice seedlings growth were analyzed, and marker genes for five different plant hormones in rice seedlings roots under the above treatments were detected by qRT-PCR in the study. The results showed that SNP can obviously inhibit the root length and plant height of the two rice seedlings, which implement mainly through its photolysis products K₄Fe(CN)₆. Moreover, the expression of the marker genes of auxin, cytokinins, abscisic acid and gibberellic acid were inhibited after the treatment of SNP and K₄Fe(CN)₆ in rice roots, but the marker gene of nitric oxide was inhibited by SNP, induced by K₄Fe(CN)₆.

Keywords: sodium nitroprusside; rice; seedling; plant hormone; gene expression

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